The molecular mechanism of entrainment of the plant circadian clock by light

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At the core of the eukaryotic circadian network, clock genes/proteins form multiple transcriptional/translational negative feed-back loops and generate a basic ~24h oscillation, which provides daily regulation for a wide range of processes. This temporal organization enhances the fitness of the organism only if it corresponds to the natural day/night cycles. Light is the most effective signal in synchronizing the oscillator to environmental cycles. Light signals mediated by photoreceptors are forwarded to the oscillator and cause an acute change in the level/activity of certain clock components that eventually results in a phase shift of the oscillation (Devlin and Kay 2001). Our aim is to reveal the molecular details of this process (also called entrainment or resetting) in *Arabidopsis thaliana*.

The plant circadian oscillator is supposed to consist of three inter-locked feedback loops Locke et al. 2006). In the first loop the morningexpressed CIRCADIAN CLOCK ASSOCIATED 1 (CCA1)/LATE ELONGATED HYPOCOTYL (LHY) transcription factors inhibit the expression of the TIMING OF CAB EXPRESSION 1 (TOC1) gene; conversely, the evening-expressed TOC1 positively regulates the transcription of CCA1/LHY. In the second loop GIGANTEA (GI) induces TOC1 expression during the afternoon/evening, while TOC1 represses GI during the night. Recent data suggested the operation of a third loop, where CCA1/LHY up-regulate the PSEUDO RESPONSE REGULATOR 7/9 (PRR7/9) genes (homologs of TOC1) in the morning and PRR7/9 proteins down-regulate CCA1/LHY expression during the day. In Arabidopsis, CCA1/LHY, GI and PRR9 may represent the primary targets of resetting light signals, merely based on the fact that these clock genes are acutely light-inducible. However, the role of their light induction in phase resetting has not been tested directly.

The primary elements of the plant light input pathway are the red/far-red light absorbing phytochromes (PHYA, B, D, E) and blue light absorbing cryptochromes (CRY1,2; Devlin and Kay 2000). However, the molecular links between phytochrome signalling and the core clock components are still missing.

In the first set of experiments we studied the function of the red/far-red absorbing phytochrome B (PHYB) photoreceptor in the resetting process. Our data show that the *phyb-9* mutation affects different parameters (phase and/or period) of rhythmic expression of components of the multi-loop circadian oscillator in Arabidopsis. This could be explained by decoupling of the different loops of the oscillator in *phyb-9*. However, we showed that genetic manipulation of a single loop has the same effect on the other loops in wild type and *phyb-9*, which indicates that absence of PHYB does not separate the individual loops. Rather, our data suggest the existence of tissue-specific clocks, which are regulated by PHYB in different ways.

The circadian oscillator responds with characteristic phase shifts to short light pulses. In the second set of experiments we investigated the effect of light induction of certain clock components on the magnitude of such phase shifts. Our data showed that the pattern of light induction of a single clock component does not correspond directly to particular phase responses, but high level of CCA1 and GI induction coincides with strong phase delays, while high level of GI and PRR9 induction coincides with strong phase advances. Phase response curves in single clock mutants indicate that among the light inducible clock components, CCA1, LHY and GI are negative elements of resetting during the subjective night, but PRR9 is a positive element of resetting during the subjective day.

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Study of the genetic relationships of oribatid mites (Acari, Oribatida) using nucleotide sequences

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Oribatid mites play important role worldwide in soil life, due to their high abundance and soil dwelling lifestyle. They are the most species rich order in the subclassis of Acari. Members of different groups of oribatid mites show very different and peculiar morphological appearance, however they belong to single taxonomical unit. There are almost 10 000 described species on the world, from which 523 species are reported in the area of Hungary to date. The species living in Hungary represent 5 major taxonomical units, divided in 77 families and 191 genera. The system of oribatid mites based on morphological features.

In order to study genetic relationships at large and small scale taxonomical levels we chose molecular markers. We used the 160 base pair long part of the 28S D3 nuclear ribosomal DNA coding domain and the 680 base pair long partial sequence of mitochondrial cytochromeoxydase I (Cox1) subunit. 96 species were collected and determined from different localities of Hungary. 36 species, representing the main