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Effect of the waste-water of sugar-works on
natural history of the river Zagyva

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Abstract

The present paper is analysing the chemical composition and natural history of the river Zagyva, as well as their change as a result of various waste-waters. It is establishing that the effect of the standing pollution depends upon the water output, while the influence of periodical pollutions /waste-water waves/ depends first of all upon the storage time of waste-water.

About the natural history of the river Zagyva there have been issued so far but a few scientific publications, only the monographs of S z e m e s /1940, 1947, 1948/ and U h e r k o v i c h /1966 a, 1968/ being known. Water-chemical data were published by S c h i k /1933/ and P a p p /1961/. The chemical and recently the biological state of its water is followed with attention systematically by organs of water conservancy, owing to the practical importance of the question / G u l y á s 1964, H a m a r 1967/. The utilization of water of the 180 km long Zagyva is manifold; irrigation, industrial water drawing, well-drain and inland water drainage. Its water output is changing /310-0,2 m³/sec./, depending mostly on the water output of streams flowing into it. The industrial waste-waters are the most important ones of the permanent sources of pollution. The galvanizing, oleiferous, tar-waters of the industrial works in Salgotrján, Hatvan and Jászbereny are spelling a grave loading on the Zagyva. /Fig.1/, mainly because of being toxic and hardly dissociable. The seasonal industrial pollutions are caused by the waste-water of the sugar-works in Selyp and Hatvan. The effect of the waste-waters pouring into it continually depends, in fact, on the water output of the Zagyva, and the waste-waters pouring into it seasonally are superposed upon that basic loading.

Materials and methods

Our laboratory performs water-quality investigations in five sample areas a month, investigating the whole polluted section in the time of waste-water waves. The analysis was carried out by means of samples obtained by drawing and netting. The chemical investigations were carried out whith COMECON-methods, the basic biological qualification with P a n t l e B u c k's /1955/ method / $S = \frac{E_{s.h.}}{\Sigma h} /$.

The figure of frequency /1-3-5/ was established by estimation. It is demonstrated by practice that P a n t l e - R u c k's formula can generally be used for detecting pollutions in case of river Zagyva. For determining Protozoa, mainly we have applied vital staining. The determination of organisms was carried out from the fundamental works of H o r t o b á g y i, H u b e r - P e s t a l o z z i, K a h l, P a s c h e r, S l a d e c k and U h e r k o v i c h.

Results

The water of the river Zagyva is of medium hardness, the dominance of ions Ca^{++} , Mg^{++} , SO_4^- , HCO_3^- is characteristic of it, the content of the total dissolved matter in it is changing /400-1300 mg/l/. Its chemical composition is influenced by its state of pollution /Table I/. It is characteristic of the state of water quality in the Zagyva, besides the waste-water waves, that it is made poly- α -mezosaprobical by the stream Tarján flowing into it at its upper reaches and carrying the industrial and house waste-waters of Salgatótarján. But in the most polluted state the water was antisaprobical. The β -mezosaprobical water of the Zagyva reaches above Hatvan is, as a result of the waste-waters of the town, again polluted /polysaprobical/. Until Jászbérény it becomes considerably clearer / β -mezosaprobical/, the waste-waters of the town exert their effect but on a short section. On the lower reaches between Jászhérény and Szolnok there is no major waste-water influence. We found the biologically clearest picture at inundations. The biological state of the lower reaches is influenced strongly by the damming effect of the river Tisza. In case of low water the pollution is stronger /Table III, 8/.

From the microorganisms the algae are dominating, mainly the diatoms whose composition changes depending upon the character of reaches and the degree of pollution /Table II, III/. Chlorophyceae appear in higher number in the vegetation period, Euglenophyta at a higher degree of pollution.

According to our observations, Cyanophyta increase at low water, in warm weather, mainly those indicating a strong pollution /Table III, 8/.

The state of water quality of the river Zagyva is the most homogeneous in the season of waste-water waves. The storage tank of the sugar-works in Selyp had a capacity of 6500 m³ in 1965, 168.000 m³ in 1967, the former one with a storing time of 8-10 days, the latter with that of 20-21 days. The waste-waters of a short storing period like that are at the beginning of the processes of anaerobic dissolution. They contain much organic matter and floating sediment, their BOD. is 90-400 mg/l. Apart from bacteria - among them streptococci - only flagellatae can be found in them./Table II, 2/ The amount of the waste-water emitted is about 1 m³/sec., corresponding to the water output of the Zagyva. Later on, that water becomes diluted but its quality is hardly changing. At that time, the water of the Zagyva becomes stinking, of black colour, strongly reductive /Table I, 3/.

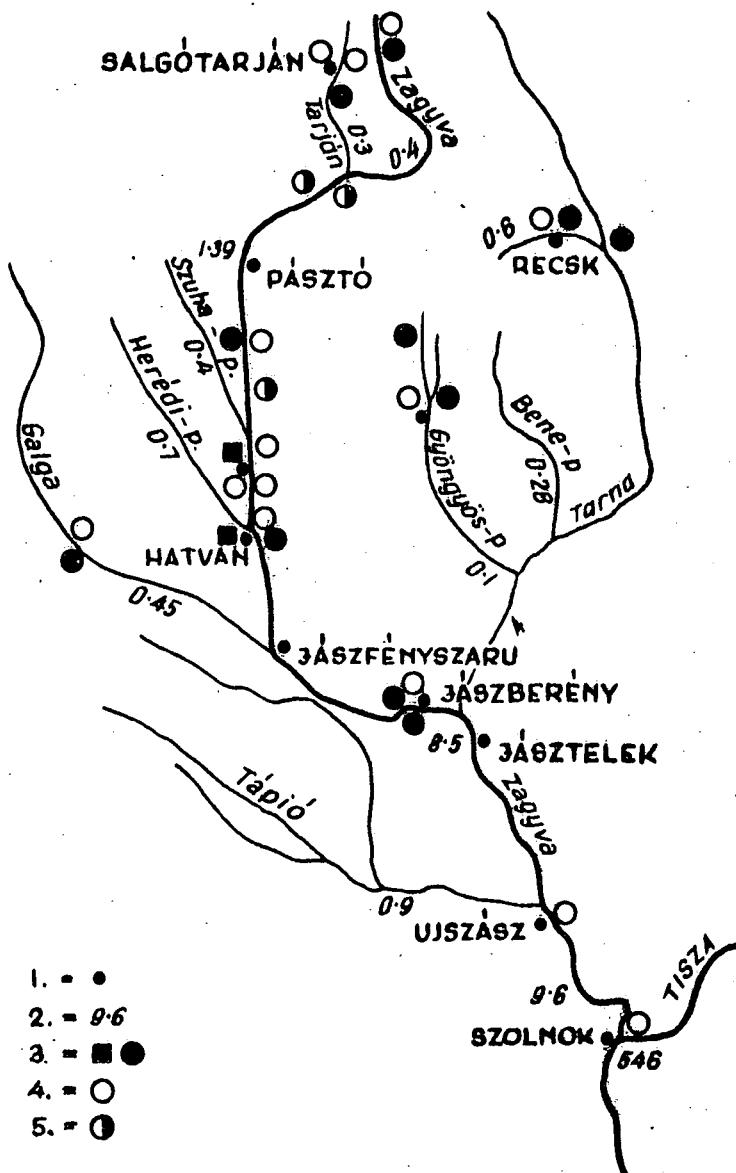


Fig. 1. Waste-water sources of the river Zagyva

Legend; 1. Settlement , 2. Mean water output ,
3. Industrial waste-water , 4. House
waste-water , 5. Industrial and house
waste-water.

The biological qualification has demonstrated only little change in the longitudinal section of the river Zagyyva. *Nitzschia palea* that appears in the lower reaches is demonstrating that some change took place in the quality of water /Table II, 3-7/. At the mouth of river, the effect of the waste-water wave can be observed well /Fig. 2/. The pollution of that degree turns the Zagyyva into a sewer, extirminating almost fully its natural fauna and flora and making its influence feel even after the waste-water wave had passed. According to the data of fishery, in campaign of sugar-works each, there perish about 2-300 q /cca. 4-600 cwt/ fish, for the most part carp, bream, and catfish.

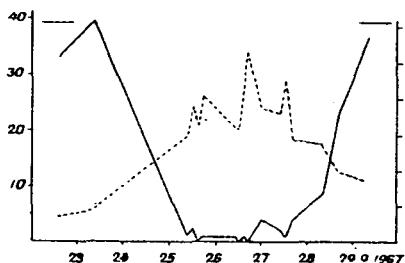


Fig. 2. Change in the oxygen management at mouth of the river Zagyyva, as a result of waste-water from the sugar-works in Selyp.

Legend; — dissolved O₂ mg/l, - - - O₂ decrease mg/l

Things are not the same in case of the sugar-works in Hatvan. This factory has a storage tank of 550.000 m³. It pours its waste-water into the river Zagyyva after finishing the period of its campaign, at high water. The waste-water itself, too, is in a more advanced state of self-purification than that of the sugar-works in Selyp. In addition to *Bacteria*, there are in its water *Protozoa* and *Algae*, as well. Owing to the stronger dilution, this waste-water can become clear in the Zagyyva, being indicated there by the discolouration of the water, too. It rather increases the nutrient content of the river and passes without any major damage /Table III, 4-6/.

A specialist, after surveying the list of taxons, can ascertain how variegated the microflora and fauna of the river Zagyyva /between Selyp and Szolnok/ is. It will be the task of further investigations to reveal the causes and regularities of the changes in the natural history.

The microorganisms of the river Zagyyva /Selyp - Szolnok/;

Schizomyctes

1. *Beggiatoa alba* Tre e v.
2. *Coccus*
- 3.

3. *Filamentous bac.*, in the time of the waste-water waves of the sugar-works and canning factory there was in the water a large amount of characteristic, long / 100-200 μ / bacteria.
4. *Sarcina palludosa* Schröeter
5. *Spaerotilus natans* Kütz., being observed in the neighbourhood of sewage pipes of the sugar-works and at low water near to the mouth, as well /Table II, 8/, with thalluses both in the plankton and benthos equally.
6. *Spirillum undulans* Ehr.
7. *Spirillum* spp.
8. *Streptococcus margaritaceus* Schröeter
9. *Thiotrix nivea* Rab. / Winog.
10. *Thiovulum* sp.
11. *Zoogloea ramigera* Itzig.

Cyanophyta

12. *Achronema articulatum* Skuja, its trichom being straight or somewhat curved, the cells strongly laced. The granules of protoplasm are denser in the longitudinal axis of the cell, in the middle of it. Cell sizes; 8,3 - 10 x 2,5 μ / Fig. 3, 13/.
13. *Coelosphaerium kutzlingianum* Nag.
14. *Coelosphaerium pusillum* van Gool
15. *Merismopedia elegans* A. Br.
16. *Merismopedia minima* G. Bock
17. *Merismopedia tenuissima* Lemm
18. *Microcystis aeruginosa* Kütz
19. *Oscillatoria amphibia* Ag.
20. *Oscillatoria chlaybea* Mert.
21. *Oscillatoria chlorina* Kütz.
22. *Oscillatoria granulata* Gard.
23. *Oscillatoria lauterbornii* Ag., the trichom of green colour is straight or mildly curved, the 2,5 μ wide and 3,3 - 5 μ long cells at the cross-wall are not laced. In the middle of cell 1-3 gas vacuoles of irregular shape can be observed /Fig. 3, 8/. It can be found in a highly polluted water, first of all associated with *Beggiatoa alba* and *Oscillatoria chlorina*.
24. *Oscillatoria limosa* Ag.

25. *Oscillatoria pseudogeminata* G. Schmid
 26. *Oscillatoria putrida* Schmidle
 27. *Oscillatoria tenuis* Ag.

Chrysophyta - *Chrysophyceae*

28. *Dinobryon divergens* Imhof

29. *Synura uvella* Ehrl

Chrysophyta - *Bacillariophyceae*

30. *Achnantes lanceolata* / Breb. / Grun.

31. *Achnantes microcephala* Kütz.

32. *Achnantes minutissima* Kütz.

33. *Amphora ovalis* Kütz.

34. *Caloneis amphibiaena* / Bory / Cl.

35. *Cocconeis placentula* Ehrl.

36. *Cocconeis placentula* var *euglypta* / Ehrl. / Cl.

37. *Cymatopleura elliptica* / Bréb. / W. Smith

38. *Cymatopleura solea* / Bréb. / W. Smith

39. *Cymatopleura solea* var. *regula* / Ehrl. / Grun

40. *Cymbella ventricosa* Kütz.

41. *Diatoma elongatum* / Lyngb. / Ag.

42. *Diatoma vulgare* Bory

43. *Fragillaria capucina* Desm.

44. *Fragillaria crotonensis* Kitt.

45. *Gomphonema constrictum* Kütz.

46. *Gomphonema olivaceum* / Lyngb. / Kütz.

47. *Gomphonema parvulum* / Kütz. / Grun.

48. *Gyrosigma attenuatum* / Kütz. / Rabh.

49. *Hantzschia amphioxys* / Ehrl. / Grun.

50. *Melosira granulata* / Ehrl. / Ralfs.

51. *Melosira granulata* var. *angustissima* / O. Müll / Hust.

52. *Melosiora varians* Ag.

53. *Navicula cryptocephala* Kütz.

54. *Navicula hungarica* Grun.

55. *Navicula pygmaea* Kütz.
 56. *Navicula rhynchocephala* Kütz.
 57. *Navicula viridula* Kütz.
 58. *Nitzschia acicularis* W. Smith.
 59. *Nitzschia capitellata* Hust.
 60. *Nitzschia closterium* / Ehr./ W. Sm.
 61. *Nitzschia gracilis* Hantzsch.
 62. *Nitzschia linealis* W. Smith.
 63. *Nitzschia palea* / Kütz / W. Smith.
 64. *Nitzschia sigmaeidea* / Ehr./ W. Smith.
 65. *Nitzschia tryblioella* Hantzsch.
 66. *Rhoicosphenia curvata* / Kütz ./ Grun.
 67. *Stephanodiscus hantzschii* Grun.
 68. *Suriella robusta* var. *splendida* / Ehr./ V. H.
 69. *Suriella ovata* Kütz./
 70. *Suriella ovalis* Bréb.
 71. *Synedra acus* Kütz.
 72. *Synedra affinis* Kütz.
 73. *Synedra ulna* / Nitzs./ Ehr.

Chrysophyta - Xanthophyceae

74. *Tribonema vulgare* Pisch., caused water-colouration in the protruded branches of the river; in its bed I did not find it /July 11th 1969/.

Euglenophyta

75. *Anisonema acinus* Duj.
 76. *Astasia klebsii* Lemm.
 77. *Euglena acus* Ehr.
 78. *Euglena intermedia* Schmitz.
 79. *Euglena polymorpha* Bang.
 80. *Euglena proxima* Bang.
 81. *Euglena oxyuris* f. *minor* Defl.
 82. *Euglena tripteris* / Duj./ Klebs.

83. *Euglena viridis* Ehr.
84. *Lepocycnelis ovum* /Ehr./ Mink.
85. *Phacus curvicauda* Svir.
86. *Phacus longicauda* /Ehr./ Duj.
87. *Phacus pleuronectes* /O. F. M./ Duj.
88. *Phacus wetsteinii* Drez.
89. *Peranema trichophorum* Chen.
90. *Menoidium falcatum* Zachar.
91. *Trachaelomonas acuminata* /Schmarck/ Stein
92. *Trachaelomonas fluviatilis* Lemm.
93. *Trachaelomonas granulosa* Playf.
94. *Trachaelomonas hispida* /Perty/ Stein
95. *Trachaelomonas intermedia* Dang.
96. *Trachaelomonas scabra* Playf.
97. *Trachaelomonas volvocina* Ehr.
98. *Trachaelomonas volvocina* var. *granulosa* Playf.

Chlorophyta - Chlorophyceae

99. *Actinastrum hantzschii* Lagerh.
100. *Ankistrodesmus falcatus* var. *acicularis* /A.Br./ West
101. *Ankistrodesmus falcatus* var. *mirabile* W. et. W.
102. *Ankistrodesmus falcatus* var. *spirilliformis* G.S. West
103. *Ankistrodesmus longissimus* Wille
104. *Ankistrodesmus setigerus* /Schoerell/ G.S. West
105. *Chlamydomonas ehrenbergii* Gor.
106. *Chlamydomonas pertusa* Chod.
107. *Chlamydomonas simplex* Pascher.
108. *Chlorella vulgaris* Bcij.
109. *Chodatella balatonica* Scherffel
110. *Chodatella ciliata* Lemm., on the poles of the ellipsoidal cell thin, curved spikes are sitting. Size of cell; 11,6 x 8,3 μ the length of spikes being 9,2 - 10,8 μ /Fig. 3,2/.
111. *Chodatella longiseta* Lemm., the length of cell is 10,8 μ , its width 8,3 μ , the length of curved spikes being 33-37,5 μ /Fig. 3,1/

112. *Chodatella quadriseta* L e m m .
113. *Cladophora glomerata* K ü t z ., a plant characteristic of the benthos and perifiton of the river Zagyva. It has a wide ecological valence being present, according to S l á d e c e k /1962/, from the clear /oligo-saprobical/ waters to the polluted /mesosaprobical/ ones. Chud y b a /1965/ describes them from a river and streams of clear water /1968/, distinguishing two characteristic groups; *Cladophora glomerata rheobenticum* occurs in flowing places, while *Cladophora glomerata limnobenthicum* in places without any flow. F j e r - d i n g s t a d /1950, 1967/ is characterizing with the *Cladophora* community a saprobical zone. Being a sessile organism, epifitic community develop on it - mainly diatom - described Chud y b a /1968/ under the name of *Cladophoretum glomeratae epiphytosum rheobenthicum* Chud y b a . An estimation of the *Cladophora* community in the river Zagyva would be reasonable even from saprobiological point of view. According to my observations, the change in pollution is well-indicated by the epifitic community on the *Cladophora*. At the passing of waste-water waves, on the *Cladophora*, *Beggiatoa alba* and *Zoogloea ramigera* settled and the frequency of *Nitzschia palea* a *Nitzschia acicularis* increased. As the pollution had ceased being, the original community reappeared.
114. *Coelastrum microporum* N ä g .
115. *Crucigenia quadrata* var. *octogona* S c h m i d l e .
116. *Crucigenia rectangularis* G a y .
117. *Crucigenia tetradedia* / K i r c h . / . W. et W.
118. *Dictyosphaerium elegans* B a c h .
119. *Dictyosphaerium pulchellum* W o o d .
120. *Eudorina elegans* E h r .
121. *Gonium pectorale* E h r .
122. *Gonium sociale* W a r m .
123. *Lagermeimia wratislaviensis* S c h o e r e d , the spikes of widened basis found on the poles and on both sides of the ellipsoidal cell lie in one level. Cell size; 6-8 x 4-5 μ , the spikes being 13-15 μ long /Fig. 3, 12/.

124. *Lambertia ocellata* var. *maxima* U h e r k o v . , the cell of size.
 140 x 14 μ is elongated spindle-shaped, narrowing
 at both ends /Fig. 3, 18/.
125. *Micraeclinium pusillum* F r e s .
126. *Oocystis lacustris* Ch o d .
127. *Oocystis crassa* var. *marssonii* P r i n t z
128. *Pandorina morum* / M a l l / B o r y .
129. *Pediastrum boryanum* M e y e n
130. *Pediastrum duplex* M e y e n
131. *Pediastrum duplex* f. *setigera* K o r s c h . , on the peaks of the outer
 cells there are sitting two-three thin spikes of 5-14 μ
 length /Fig. 3, 16/
132. *Scenedesmus acuminatus* / L a g e r h ./ Ch o d .
133. *Scenedesmus acutus* M e y e n
134. *Scenedesmus anomalous* / G. M. S m i t h / T i f f . , cell size;
 6-8 x 1,7 μ , spike 7,5-10 μ /Fig. 3, 4-5/.
135. *Scenedesmus anomalous* / G. M. S m i t h / T i f f . varians? , the cells
 forming coenobium in pairs, bending outwards. Size
 cell; 10 x 1,7 μ , spike 9-10 μ /Fig. 3, 3/.
136. *Scenedesmus anomalous* / G. M. S m i t h / T i f f . forma? on the ends of
 cells of size; 8,3 x 1,7 μ a short spike of 3-3,5 μ
 is sitting /Fig. 3, 6/.
137. *Scenedesmus bicaudatus* / H a n g s ./ Ch o d .
138. *Scenedesmus denticulatus* L a g e r h .
139. *Scenedesmus ecornis* / R a l f s ./ Ch o d .
140. *Scenedesmus ellipsoideus* Ch o d .
141. *Scenedesmus granulatus* W. et W.
142. *Scenedesmus intermedius* Ch o d .
143. *Scenedesmus intermedius* var. *balatonicus* H o r t o b .
144. *Scenedesmus intermedius* var. *bicaudatus* H o r t o b .
145. *Scenedesmus nanus* Ch o d . , cell size; 7,5 x 5 μ , spike
 /Fig. 3, 10/.
146. *Scenedesmus opoliensis* P. R i c h t .
147. *Scenedesmus protuberans* F r i t s c h .
148. *Scenedesmus quadricauda* / T u r p ./ B r é b .
149. *Scenedesmus quadricauda* var. *setosus* K i r c h . /Syn; Sc. *longus*
 M e y e n , Sc. *longus* var. *nägeli* / B r é b . /

G. M. Smith, *Sc. nägelii* Bréb., the coenobium consists of 4-8 cells, the cells being spindle-shaped, their ends bluntly peaked or rounded. On the poles of inner cells a straight or somewhat curved spike of changing length is sitting. Cell size: 6,7-10 x 2,5-5 μ , the length of spikes being 5-10 μ . The specimens found by me /Fig. 3, 9, 11, 14, 15/ can be classed into the form-circle of *Sc. quadricauda* var. *setosus* Kirch., if we consider the taxons of *Sc. longus*, *Sc. longus* var. *nägelii*, *Sc. nägelii* - those their specimens where the inner cells are spiky, as well, - to be synonymous.

150. *Sc. spinosus* Chod.

151. *Selenastrum gracile* Reinsch

152. *Sphaerellopsis gleocystiformis* /Dill/ Gerloff, Cell size; 12 x 9 μ together with cell-membrane; 20 x 18, the flagellum being 21 μ long /Fig. 3, 17/.

153. *Spondylomorium quadrernarium* Ehr.

154. *Tetraedron minimum* /A. Br./ Hansg.

155. *Tetraedron muticum* /A. Br./ Hansg.

156. *Tetraedron glabrum* /Röhl/ Ahl. et Tiff.

157. *Tetrastrum staurogenieforme* /Schoered/ Lemm.

Chlorophyta - Conjugatophyceae

158. *Closterium acerosum* Ehr.

159. *Closterium strigosum* Bréb.

160. *Spirogyra* sp.

161. *Staurastrum paradoxum* Meyen

Zooflagellata

162. *Bodo globosus* Stein

163. *Bodo mutabilis* Klebs

164. *Bodo putrinus* Lemm.

165. *Bodo repens* Klebs.

166. *Bodo saltans* Ehr.

167. *Cercobodo agilis* /Moroff/ Lemm.

168. *Cercobodo longicauda* /Stein/ Senn

169. *Desmarella moniliiformis* Kent.

170. *Hexamitus inflatus* Du j.
 171. *Hexamitus pusillus* K le b s .
 172. *Monas vulgaris* /C i e n./ S e n n .
 173. *Oicomonas sociabilis* K e n t
 174. *Tetramitus pyriformis* K I e b s
 175. *Trepomonas agilis* Du j.
 176. *Trepomonas rotans* K l e b s
 177. *Trepomonas steinii* K l e b s
 178. *Trigonomonas compressa* K l e b s

R h i s o p o d a

179. *Actinophris sol* E h r .
 180. *Actinosphaerium echicornii* E h r .
 181. *Amoeba radiosa* D u j .
 182. *Arcella vulgaris* E h r .

C i l i a t a

183. *Aspidisca costata* C l . et L a c h .
 184. *Carhesium* sp.
 185. *Chilodonella cucullulus* O.F. M ü l l .
 186. *Chilodonella uncinata* E h r .
 187. *Chinetochilum margaritaceum* P e t r y .
 188. *Coleps hirtus* N i t z s .
 189. *Colpidium colpoda* S t e i n
 190. *Cyclidium citrulus* C h o n
 191. *Dileptus anser.* O. F. M ü l l .
 192. *Euplotes charon* S t e i n
 193. *Glaucoma scintillans* E h r .
 194. *Halteria grandinella* E h r .
 195. *Holophrya nigricans* L a u t e r .
 196. *Lionotus fascicola* E h r .
 197. *Loxophyllum helus* S t o k .
 198. *Metopus es* C l . et L o c h .
 199. *Metopus contortus* L e v . ?
 200. *Oxytricha fallax* S t e i n ?
 201. *Paramecium* sp.
 202. *Pleuronema crassum* D u j . ?
 203. *Plagiophylla nasuta* S t e i n
 204. *Prorodon terres* E h r .
 205. *Spirostomum ambiguum* E h r .
 206. *Stentor polymorphus* E h r . - S t e i n
 207. *Stylochichlia mytilus* E h r . ?
 208. *Tachiosoma pellionella* M ü l l . - S t e i n

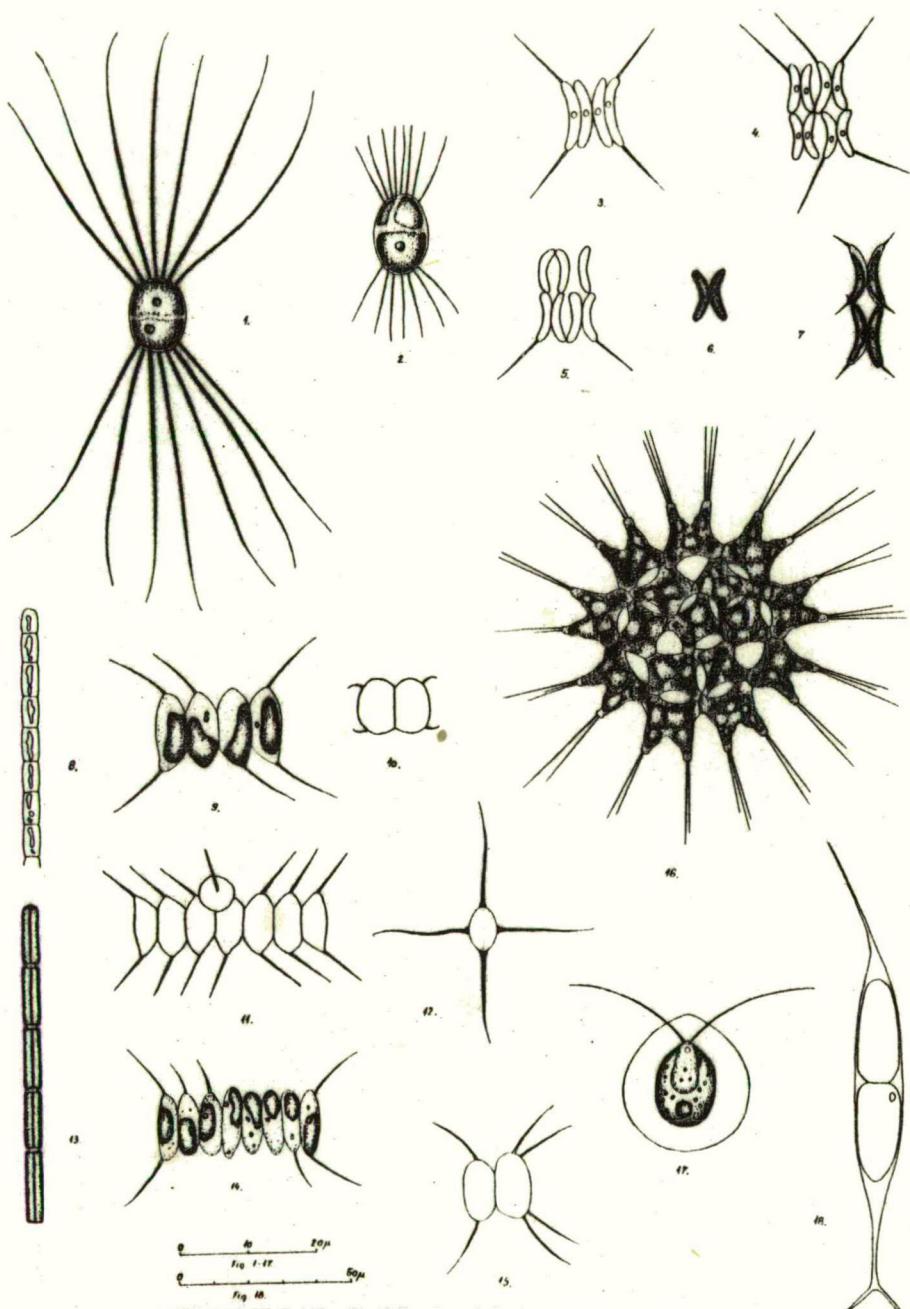


Fig. 3. 1. *Chodatella longiseta*, 2. *Chodatella ciliata*, 3. *Scenedesmus anomalus forma*, 4.5. *Scenedesmus anomalus*, 6. *Scenedesmus anomalus* var. *acaudatus*, 7. *Scenedesmus anomalus forma*, 8. *Oscillatoriella lauterbornii*, 9.11.14.15. *Scenedesmus quadricauda* var. *setosus*, 10. *Scenedesmus nanus*, 12. *Lagerheimia wratislawiensis*, 13. *Achronema articulatum*, 16. *Pediastrum duplex* f. *setigera*, 17. *Sphaerellopsis gleocystiformis*, 18. *Lambertia ocellata* var. *maxima*

209. *Urostyla* sp.
 210. *Uronema marinum* Du j.
 211. *Vorticella campanula* Ehr.
 212. *Vorticella convallaria* Noland

Summary

The defence of our natural water supply belongs to our first-class tasks. By increasing the sources of pollution, the ability of purification in our waters decreases. That is meaning a problem, mainly in case of receivers of a small and changing water output like the river Zagyva is. The change in the degree of pollution engendered by the several and continuous waste-water intakes depends first of all on the water output of the river Zagyva. The major pollutions are caused by the seasonal waste-water disposals of the sugar-works. With chemical and biological investigations we have followed with attention the effect of the waste-water of the sugar-works on the Zagyva and found a considerable difference between the effects of the waste-water waves of the two sugar-works, caused by the different storing periods. We have referred with a few representative examples to the chemical and biological state of the river Zagyva.

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	pH	Alkal. w°	Ca ⁺⁺ mg/l	Mg ⁺⁺ mg/l	Na ⁺ mg/l	K ⁺ mg/l	Cl ⁻ mg/l	SO ₄ ⁻⁻ mg/l	HCO ₃ ⁻⁻ mg/l	PO ₄ ⁻⁻⁻ mg/l	NH ₄ ⁺ mg/l	NO ₂ ⁻ mg/l	NO ₃ ⁻ mg/l	O ₂ decrease mg/l	Diss. O ₂ mg/l	BOD 5 days mg/l	CO ₃ ⁻⁻ mg/l	Free CO ₂ mg/l	H ₂ S mg/l	Bio ₁ .
1. Pure	7,3	7,2	107	51,1	71,5	14,7	55,0	209	439	0,67	∅	0,03	10,3	2,04	14,0	5,92	∅	0,96	∅	8-a
2. Pol- luted	8,5	7,6	93,8	84,1	76,0	34,0	71,1	265	463	2,00	0,3	∅	6,3	5,24	7,36	8,74	24,0	∅	∅	p
3. Strong- ly pol- luted	8,1	11	112	52,0	90,0	20,5	46,1	129	671	0,67	5,1	0,03	0,76	62,4	∅	116,5	6,00	∅	∅	a

Table 1. Various pollution of the river Zagyva

Legend: 1. Jászfényszaru Apr. 24 1968
 2. Jászfényszaru Apr. 30 1968
 3. Jászfényszaru Sept. 20 1967

Table 2. Biological evaluation of the waste-water wave from the sugar-works in Selyp

Table 3. Biological evaluation of the waste-water wave from the sugar-works in Hatvan.

Sample areas
April 30th 1968